

PROCESSING MICROARRAYS

I. REHYDRATION (remember to *etch* arrays on back (label side down) before processing)

- a) pour 42°C 1x SSC into a humidifying chamber
- 100ml solution: 95ml water
 5ml 20x SSC
- b) place array upside down (label side down) in chamber ~60 sec until all spots look rehydrated (spots will start to “glisten”)
- c) dry slide on 100°C heat block for ~2 sec

II. SHAMPOO (helps to reduce background signal)

- a) soak slides in 65°C 3x SSC + 0.2% SDS, 5 min
- b) using hyb oven & glass slide dish
- 500ml solution: 75ml 20x SSC
 10ml 10% SDS
 water to fill
- c) transfer slides to fresh slide rack (minimizes SDS carryover) rinse with water ~ 30” @ RT
- d) rinse in 95% EtOH
- e) spin dry ~ 5’ (400k)

III. BLOCKING

- a) measure 335ml of 1-methyl-2-pyrrolidinone (clear and not yellow) into a clean, dry 500ml beaker & dissolve 5.5g of succinic anhydride
- b) *immediately* add 15ml of 1M sodium borate pH 8.0
- c) *quickly* pour blocking solution into a glass dish & plunge slides rapidly ~30”
- d) put lid on box and put on rotator ~15’
- e) boil enough water (~95°F) in glass dish (microwave) & swish slide rack for a few seconds, let incubate ~1’
- f) *quickly* transfer slide rack to glass dish of 95% EtOH & plunge for a few seconds
- g) spin slides ~1’ (400k)
- h) store slides in plastic slide box until ready to be used (2 weeks max storage)